#### Form Approved REPORT DOCUMENTATION PAGE OMB No. 074-0188 Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, includ suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503 1. AGENCY USE ONLY 3. REPORT TYPE AND DATES COVERED 2. REPORT DATE (Leave blank) 5/1/98 Final Technical 2/1/97-1/31/98 4. TITLE AND SUBTITLE 5. FUNDING NUMBERS Genetic Analysis of Vertbrate Circadian Rhythmicity F49620-97-1-0048 2312/CS 61102F 6. AUTHOR(S) Gregory M. Cahill, Ph.D. 7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) AFRL-SR-BL-TR-98-University of Houston 053x 4800 Calhoun Road Houston TX 77204 9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) 10. SPONSORING / MONITORING AGENCY REPORT NUMBER Air Force Office of Scientific Research/NL 110 Duncan Ave 19980722 036 Bolling AFB, DC 20332 11. SUPPLEMENTARY NOTES 12b. DISTRIBUTION 12a. DISTRIBUTION / AVAILABILITY STATEMENT CODE Approved for public roleage; distribution unlimited. 13. ABSTRACT (Maximum 200 Words) The goal of this project was to develop technology needed to identify vertebrate circadian clock genes. An understanding of basic molecular mechanisms of biological clocks is important in designing treatments of the performance deficits, sleep disorders and other problems associated with jet lag, shift work, and organic circadian clock disorders in humans. A novel automated image analysis system was developed to measure circadian locomotor rhythms. This system was used to measure behavioral rhythms in zebrafish, a vertebrate organism that is useful for large scale mutagenesis screens. A screen for dominant mutations that alter the period of zebrafish locomotor rhythms was initiated, and 1500 animals with different mutagenized genomes were tested. The progeny of 40 putative mutants are being tested to determine whether they carry authentic clock mutations. 14. SUBJECT TERMS 15. NUMBER OF PAGES 21 Circadian rhythms, Genetics 16. PRICE CODE 17. SECURITY 18. SECURITY 19. SECURITY 20. LIMITATION OF **CLASSIFICATION** CLASSIFICATION **CLASSIFICATION ABSTRACT**

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OF ABSTRACT

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## Summary

The goal of this project was to identify vertebrate circadian clock genes through mutational analysis of zebrafish circadian rhythm. The original proposal was for three years, and the specific aims included: 1) development of an automated image analysis method for measurement of locomotor activity rhythms; 2) a screen of 5,000 mutagenized individuals for dominant mutations; 3) genetic mapping of confirmed clock mutations; 4) characterization of mutant phenotypes, and 5) determination of the effects of visual system mutants on circadian rhythms. In the one year that was funded by the AFOSR, we accomplished the first specific aim and initiated the second specific aim. Specific aims 3-5 will be pursued with funding from other sources.

#### **Publications**

Bégay, V., Falcón, J., Cahill, G.M., Klein, D.C. and Coon.S.L. 1998. Transcripts Encoding Two Melatonin Synthesis Enzymes in the Teleost Pineal Organ: Circadian Regulation in Pike and Zebrafish but not in Trout. *Endocrinology*: **139**:905-912

Klein, D.C., Coon, S.L., Roseboom, P.H., Weller, J.L., Bernard, M., Gastel, J.A., Zatz, M., Iuvone, P.M., Rodriguez, I.R., Begay, V., Falcon, J., Cahill, G.M., Cassone, V.M. and Baler, R. 1997. The melatonin rhythm-generating enzyme: Molecular regulation of serotonin N-acetyltransferase in the pineal gland. *Rec. Prog. Horm. Res.* 52: 307-358

Hurd, M.W., DeBruyne, J., Straume, M. and Cahill, G.M. Circadian rhythms of locomotor activity in zebrafish. In press, *Physiology and Behavior*.

# Manuscript submitted

Cahill, G.M., Hurd, M.W. and Batchelor, M.M. Circadian rhythmicity in the locomotor activity of larval zebrafish. Submitted, *Neuroreport*..

#### **Published Abstract**

Hurd, M.W., DeBruyne, J, and Cahill, G.M. 1997 Circadian rhythms of locomotor behavior in zebrafish. *Soc. Neurosci. Abstr.* 23:1322.

## Progress on Specific Aims

Specific Aim 1: Development of an automated video image analysis for measurement of circadian behavioral rhythms in larval zebrafish. This aim has been completed and reported in a manuscript submitted to the journal *Neuroreport*. The methods and results are described below. This project entailed development of a computer macro program, entitled Swimming Macro, written in Analytical Language for Images, and run within the the Optimas image processing program. The code is included in the appendix.

#### Methods: Specific Aim 1

Animals: The zebrafish used in these studies were of an AB strain that was obtained from the University of Oregon. They were raised to 10 days of age in a 14:10 light:dark cycle at 28.5°C, and they were fed *Paramecium* through the day before activity recording began. During activity recording they were housed individually in 0.75 ml wells. They were not fed during the 130-hr recording period. All protocols were reviewed and approved by the University of Houston Institutional Animal Care and Use Committee.

Recording apparatus: Fish were housed individually in up to 100 cylindrical wells (0.75 cm diam, 1 cm deep) drilled in a 10 X 10 grid in a 1.2 cm thick specimen plate made of translucent white polyethylene. A diffuse axial illuminator (Edmund Scientific, Barrington, NJ) placed above the wells provided even, infrared (>700 nm) illumination and eliminated reflection from the water surface. The wells were backlighted by a placing a mirror beneath the specimen plate; this enhanced the contrast of the fish relative to background. Images were collected from a CCD camera with a 28 mm lens, automatic gain control and shading correction (Hamamatsu Photonics, Hamamatsu, Japan). The specimen plate, illumination system and camera head were enclosed in a refrigerated incubator maintained at  $24 \pm 0.1$ °C. Monochrome images (640 X 480 pixels, 8 bit resolution) were captured by a Flashpoint 128 video card (Integral Technologies, Indianapolis, IN) in a computer with a Windows operating system and either a Pentium 166 or Pentium Pro II 300 processor (Intel, Santa Clara, CA).

Data acquisition: Data were acquired automatically via Optimas (Seattle, WA) image analysis software, controlled by a macro written in Analytical Language for Images. The code fro this macro is inclluded in the appendix. Every 4 or 5 min, a series of 60 images was collected (1/sec) and stored in memory, then the distance that each fish moved in the series of images was measured. During set up of an experiment, the image frame was divided into rectangular cells enclosing each of the 100 wells, and a pixel value threshold that distinguished all of the fish from the lighter background was set. The coordinates of the centroid of each fish were determined for each frame of the series, and from these data the path through which each fish moved in the series of images was plotted. The length of the path of each fish was measured and stored in a text file, and the images in memory were discarded before reinitiating the cycle. The analysis of a series of 60 images of 100 fish required 2.5-4 min of processing time, depending on the microprocessor and version of Optimas that were used.

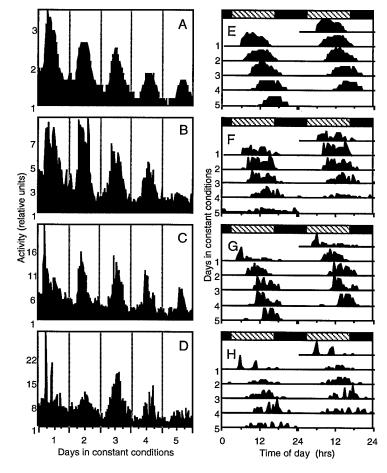
Two types of errors in locating the coordinates of the fish were encountered with this system: 1) In some cases, a fish was not recognized because too few pixels crossed threshold. When no fish was recognized, the previous location of the fish was recorded (i.e. no movement). This occurred most often when fish were inactive near the bottom of the well, so this probably did not contribute much to noise in the records. 2) In other cases, more than one object was recognized in a well, either because the fish image crossed threshold in two or more separate locations, or because a background area crossed threshold. In these cases the mean of the coordinates of all object centroids was recorded.

Experimental Design: Typically, behavioral recording began at the normal time of the light to dark transition on the tenth day post-fertilization and was continued for five to six days in constant infrared illumination. In another series of experiments, two groups of animals were exposed to opposite light cycles from five to ten days of age, and then they were monitored simultaneously in constant conditions to determine whether the recorded rhythms reflected the phase of the prior entraining light cycle.

Data analysis: Data were plotted and analyzed with Chrono II software (T. Roenneberg, University of Munich). The path lengths recorded during each 30 min period of an experiment were averaged, then plotted as a function of time. For actogram plots and periodogram analysis, a 26 h centered moving average was subtracted from the data to remove long term trends in overall activity levels. This results in positive values at times of peak activity, and negative values during troughs. In actograms, the positive values are plotted as a percent of the 26 hour mean, and negative values are omitted. Estimates of circadian period were determined by chi-square periodogram.

<u>Figure 1.</u> Circadian activity rhythms of larval zebrafish in constant conditions. A-D, Continuous records of activity. E-H, double-plotted actograms derived from the activity records in

A-D. A and E, averaged activity of 100 fish during a five day experiment. B-D and F-H are records from 3 representative individuals to illustrate the range of activity patterns observed in that experiment. The light cycle prior to the experiment is indicated by the shaded (light) and black (dark) bars above the actograms. The actograms, which plot activity levels above the daily mean, emphasize that the fish were most active during the subjective day, and that the freerunning periods of the activity rhythms are longer than 24 hr.



## Results: Specific Aim 1.

The locomotor activity of larval zebrafish in constant conditions was rhythmic with an average freerunning circadian period of 25.6 h. Examples of raw activity data and actograms for the average of a group of 100 fish and for 3 individuals are shown in Fig 1. Periodogram analysis indicated that over 95% of the animals expressed significant periodicity in the range of 23-28 hours in each of 5 experiments. The activity of every rhythmic animal was highest during the subjective day, with an approximately twofold difference between average trough and peak activity levels. A decrease in both trough and peak activity levels was observed over the course of the five day recording period in every experiment. Among experiments, average freerunning periods ranged from 25.2 to 26.1 hrs, and the within-experiment standard deviations in period ranged from 0.5 to 1.0 hours.

In order to ensure that the measured activity rhythms resulted from endogenous circadian rhythmicity, rather than some uncontrolled rhythmic variable in the environment or recording system, sibling groups of fish were raised in oppositely phased light:dark cycles, then activity of the two groups was monitored simultaneously in constant conditions. As shown in Fig. 2, the activity rhythms of the two groups of animals in constant conditions had opposite phases, with activity peaks occurring during the expected light period for each group. This demonstrates that the measured rhythms are driven by endogenous, entrainable circadian clocks, and can not be explained by extrinsic factors.

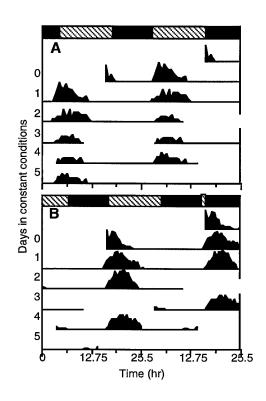
All zebrafish tested under these conditions survived the five day recording period without food or water changes. Some were removed from the wells at the end of the recording period and raised. All of these developed normally and began to breed at two to three months of age.

### Conclusion: Specific Aim 1

The locomotor activity of larval zebrafish is regulated by a light-entrainable circadian clock. The resulting activity rhythms in constant conditions are robust, with consistent freerunning periods and a consistent phase relative to the prior light cycle. Activity rhythms of 100 individuals

can be monitored simultaneously by automated video image analysis. This system should provide the efficiency and precision necessary for identification and characterization of zebrafish circadian clock mutants.

Figure 2. The phase of the zebrafish activity rhythm in constant conditions is determined by the preceding light:dark cycle. Double-plotted actograms of averaged activity from animals previously exposed to either a normal (A) or reversed (B) light:dark cycle, then monitored simultaneously (30 per group). The actograms are plotted on a 25.5 hr time scale, which approximates the freerunning periods of the rhythms. The light exposure during the 51 hours prior to placing the animals in constant conditions is indicated by the shaded bars above the graphs. The blank interval on Day 3 resulted from an 18 hr interruption of data collection.



Specific Aim 2: Mutagenesis and screen for dominant mutations in zebrafish circadian clock genes. We have made significant progress on this aim. Behavioral rhythms from approximately 1,500 mutagenized have been recorded and analyzed. Forty putative mutants have been selected on the basis of lengthened or shortened period of the circadian behavioral rhythm. The progeny of these 40 putative mutants are being tested for genetic transmission of clock mutations.

## Methods, Specific Aim 2

Mutagenesis: Male zebrafish, 3-5 months old, were immersed 3-5 times for one hour in 3 mM N-ethyl-N-nitrosourea (ENU) to mutagenize progonial sperm cells. After one month recovery, these animals were crossed with normal females. Each of the resulting progeny is expected to be heterozygous for a unique combination of mutant loci.

Screening: Progeny of mutagenized males (F1s) were tested for changes in behavioral circadian rhythms by the method described under Specific Aim 1. Behavioral rhythms were measured for 5 days, and putative mutant individuals with freerunning periods shorter than 24 hours or longer than 27 hours were selected and raised to breeding age. Progeny of these animals (F2s, 15-30 each) are then tested to determine whether the period defect segregated with a 1:1 ratio.

# Results, Specific Aim 2

To date, we have screened 1500 F1s. Forty of these animals that expressed circadian rhythms with long or short periods have been selected. The progeny of these mutants will be tested to determine whether the period changes are due to dominant mutations.

### Conclusions, Specific Aim 2

The data to date indicate that it will be possible to identify circadian clock genes by mutational analysis in zebrafish. This component of the project is continuing with funding from another source.

# Appendix:

```
Computer code for swimming macro.
Title
       Swimming.Mac
By
       Matthew M. Batchelor
       Meyer Instruments, Inc.
       Houston, TX
Support Software
       Minimize.Mac
       Optdlg.Oml
Modification Log
       08-07-97: Originally based on TstDlg.Mac - MMB
// First, prevent recursive creation of this dialog
If (IsObject ("Swimming_hWndDlg")) {
       Beep();
       If (! Prompt ("Recursive attempt to create dialog box detected!\r\n\r\nContinue?", 2))
              Pause();
       }
}
ChangeCursor (1);
                     // Make cursor into hourglass
// ----- VARIABLES used in the dialog box -----
CHAR
       Swimming_Directory = MacroPathAndName[0, ]; // Save current directory
INTEGER
       Swimming_hWndDlg; // The window handle of the new dialog box
BOOLEAN
       Swimming_IconOPTIMAS = FALSE; // Set TRUE to make OPTIMAS iconic
CHAR
       Swimming_T109 = ".
Swimming_T110 = ".
       Swimming_T118 = "
       Swimming_T141 = ",
Swimming_T142 = ",
Swimming_T143 = ",
       Swimming T144 = "1"
       Swimming_T145 = "99".
       Swimming_T146 = "3",
Swimming_T147 = "3",
       Swimming_T148 = Swimming_T144,
       Swimming_T149 = Swimming_T145,
```

```
Swimming_T161 = ".
Swimming_T162 = ".
      Swimming_T163 = "
      Swimming_T999 = ";
CHAR
      Swimming_ArithmeticOpsNames [ , ] = ";
REAL
      Swimming_ROIWide;
REAL
      Swimming_ROIHigh;
INTEGER
      Swimming_NRows;
INTEGER
      Swimming_NCols;
REAL
      Swimming_SampleWide;
REAL
      Swimming_SampleHigh;
REAL
      Swimming_ROIImage [,];
REAL
      Swimming_ROIAnalyze [ , ];
Swimming_ROIImage = ROI;
Swimming_ROIAnalyze = ROI;
REAL
      Swimming_DisplaySuperMArCentroids [ , , ] = ";
//If (!IsWindow ("Swimming_DisplaySuperMArCentroids"))
      ViewBox (Swimming_DisplaySuperMArCentroids, 0x0800);
INTEGER
      Swimming_DataNoOfBlobs [ 10, 10, 60] = ";
REAL
      Swimming_DataCentroid [ 10, 10, 60, 2] = ";
REAL
      Swimming_DataLength [ 10, 10] = ";
INTEGER
      Swimming_Row,
      Swimming Col,
      Swimming_Img;
CHAR
      Swimming_cFileName [,];
INTEGER
```

```
Swimming_cFileHandle;
// ----- Startup code -----
// Make OPTIMAS iconic ?
if (Swimming IconOPTimas)
RunMacro (PathVariable: "macros/minimize.mac");
// Load the DLL that handles custom dialog boxes
Swimming hLib = LoadMacroLibrary( "optdlg.oml");
// Register function from windows which gives system tick count in msec
// Swimming hGTC = Register ("User", "GetTickCount", "%D");
// Swimming_hGTC = Register ("Kernel32", "GetTickCount", "%D");
// ----- FUNCTIONS used in the dialog box ------
// Function to update information in the dialog box
Define Swimming_Update ()
      // Update ROI text
      Swimming_ROIWide = Swimming_ROIAnalyze [1,0] - Swimming_ROIAnalyze [0,0]
];
      Swimming_T109 =
             ToText (Swimming_ROIWide, "%.2f"):
             " (": GetOrSetField (ActiveCalibration, 103): ".)"
      Swimming ROIHigh = Swimming ROIAnalyze [0, 1] - Swimming ROIAnalyze [1, 1]
];
      Swimming_T110 =
             ToText (Swimming_ROIHigh, "%.2f"):
             " (": GetOrSetField (ActiveCalibration, 103): ".)"
      // Update row and column variables
      FromText (Swimming_T146, Swimming_NRows);
      FromText (Swimming_T147, Swimming_NCols);
      Swimming_SampleWide = Swimming_ROIWide / (Real) Swimming_NCols;
      Swimming SampleHigh = Swimming ROIHigh / (Real) Swimming NRows;
// The init macro can do any special processing such as creating viewboxes
Define Swimming_InitMacro()
      // Call update function to start everything off right
      Swimming_Update ();
// Button to prompt for path, prefix and extension
Define Swimming_B119()
      // Declare local variables
      Local Char TempPath [];
      Local Char TempFileName [,];
```

```
// Initialize TempPath
        TempPath =
               FileSplitPath (ImageFile)[0,];
        TempPath :=
               FileSplitPath (ImageFile) [1,];
       // Prompt the user for the filename
        TempFileName = GetFileName ( TempPath : "*.*", -2, -1, TRUE );
       // Get file name (with error trap)
       If (TempFileName != FALSE)
               // Open the image indicated by the user
               OpenImage (TempFileName);
               // Remember the roi used by the image
               Swimming_ROIImage = ROI;
               // Update the dialog box text
               Swimming_T141 = FileSplitPath (TempFileName) [0,];
              Swimming_T141 := FileSplitPath ( TempFileName ) [ 1 , ];
Swimming_T142 = FileSplitPath ( TempFileName ) [ 2 , ];
               Swimming_T142 = Swimming_T142 [0.3]:
               Swimming_T143 = FileSplitPath (TempFileName) [3,];
       }
// Button to load images into memory
Define Swimming_B120()
       // Declare local variables
       Local Char
              TempFileName;
       Local Char
              TempCycleC;
       Local Char
              TempImageC;
       Local Integer
              Start,
              End:
       // Initialize local variables
       FromText ( Swimming_T144 , Start);
       FromText (Swimming T145, End);
       // Clear names array
       Swimming_ArithmeticOpsNames = ";
      // Update user info
       Swimming_T118 = "Loading images into memory!";
      // Load all images from with the current prefix, cycle, and extension
      // into the arithmetic ops buffer in sequential order
```

```
For (j = Start; j \le End; j ++)
             For (i = 0; i < 99; i ++)
                    // -- Build the file name
                    // Start with the path
                    TempFileName = Swimming_T141;
                    // Add the prefix
                    TempFileName := Swimming_T142;
                    // Add the Cycle
                    If (j < 1000)
                           TempCycleC = ToText(j);
                    If (j < 100)
                           TempCycleC = "0" : ToText(i);
                    If (j < 10)
                           TempCycleC = "00" : ToText(j);
                    TempFileName := TempCycleC ;
                    // Add the Image
                    If (i < 100)
                           TempImageC = ToText(i);
                    If (i < 10)
                           TempImageC = "0": ToText(i);
                    TempFileName := TempImageC;
                    // Add the extension
                    TempFileName := Swimming_T143;
                    // If the file exists
                    If (OpenFile (TempFileName, 0x4000)!= FALSE)
                           // Put it on the arithmetic ops stack
                           FileToList (TempFileName);
                           // Remember the names on the stack
                           Swimming ArithmeticOpsNames ::=
                                  Swimming_T142 : TempCycleC : TempImageC :
Swimming T143;
             // Update user info
             Swimming_T118 = "Images from Cycle": ToText(j): "loaded into memory!";
      // Update user info
       Swimming_T118 = "Images loaded into memory!";
// Button to playback images in memory
Define Swimming_B121()
      For (i = 0; i < GetShape (Swimming ArithmeticOpsNames) [0]; i++)
```

```
StatusBar = "Arithmetic Ops Copy of ": Swimming_ArithmeticOpsNames [i,];
              SelectROI (Swimming ROIImage);
             ArithmeticOp ("Copy", Swimming_ArithmeticOpsNames [i,]);
// Button to clear images in memory
Define Swimming_B122()
      For (i = 0; i < GetShape (Swimming_ArithmeticOpsNames) [0]; i + +)
        StatusBar = "Deleting ": Swimming_ArithmeticOpsNames [i,];
              DeleteImage (Swimming ArithmeticOpsNames [i,]);
      // Clear name array
       Swimming ArithmeticOpsNames = ";
// Button to set region of interest
Define Swimming_B123()
       // Prompt user
      If ( Prompt ( "Use current ROI as Analysis ROI?", 2))
             // Remember this ROI
              Swimming_ROIAnalyze = ROI;
              Else
              // Activate ROI tool
              SelectROI();
             // Remember this ROI
             Swimming ROIAnalyze = ROI;
      // Call update function
       Swimming_Update ();
// Button to test sample layout
Define Swimming_B124()
       // Clear existing overlaid screen objects
       ClearScreen ();
       SelectROI ( Swimming_ROIAnalyze );
       // Turn off screen updated
       BeginOrEndUpdateBlock(TRUE);
       // Loop through the rows
       For (j = 0; j < Swimming_NRows; j ++)
              // Loop through the columns
```

```
For (i = 0; i < Swimming NCols; i ++)
                    // Create an area at the bounds of the ROIS
                    CreateArea
                           (ROI [0, 0] + ((Real) i * Swimming_SampleWide)):
                           ROI [0, 1] - ((Real) j * Swimming_SampleHigh))
                           ROI [0,0] + ((Real) i * Swimming_SampleWide)):
                            ROI [0,1] - ((Real) (j+1) * Swimming_SampleHigh))
                          (ROI [0,0]+((Real)(i+1)*Swimming SampleWide)):
                           (ROI[0,1]-((Real)(j+1))*Swimming SampleHigh))
                           (ROI [0,0]+((Real)(i+1)*Swimming_SampleWide)):
                           ROI [0, 1] - ((Real) j * Swimming SampleHigh))
      // Turn on updates and refresh everything
      BeginOrEndUpdateBlock(FALSE);
// Function to loop through the sample areas
Define Swimming_SampleLoop()
      // Clear existing overlaid screen objects
//
      ClearScreen ();
      // Loop through the rows
      For (j = 0; j < Swimming_NRows; j ++)
             // Loop through the columns
             For (i = 0; i < Swimming NCols; i ++)
                    // Create an ROI
                   SelectROI
                          (Swimming_ROIAnalyze [0,0]+((Real) i *
Swimming_SampleWide)):
                          (Swimming_ROIAnalyze [0,1]-((Real) j *
Swimming_SampleHigh))
```

```
(Swimming_ROIAnalyze [0,0]+((Real)(i+1)*
Swimming SampleWide)):
                           (Swimming_ROIAnalyze [0, 1] - ((Real) (j+1)*
Swimming_SampleHigh))
                           );
                    // Set global variables equal to loop counters
                    Swimming Row = i;
                    Swimming_Col = i;
                    // Run the macro to process this ROI
                    RunMacro (Swimming Directory: "Process/Process.Mac");
              }
// Go Button to process images from disk
Define Swimming_B140()
      // Declare local variables
      LOCAL INTEGER
             CycleStart, CycleEnd;
      // Initialize local variables
      FromText (Swimming_T148, CycleStart);
      FromText (Swimming_T149, CycleEnd);
      // Set up data extraction
      SetExport (Null, -2, FALSE);
      SetExport (mArCentroid, 1, TRUE);
      SetExport (ArCentroid, 1, TRUE);
      SetExport (mLnLength, 1, TRUE);
      SetExport (LnLength, 1, TRUE);
      // Initialize summary data vectors
      GLOBAL INTEGER
             Swimming_DataNoOfBlobs [ 10, 10, 60] = "";
      GLOBAL REAL
             Swimming_DataCentroid [ 10, 10, 60, 2] = "";
      GLOBAL REAL
             Swimming_DataLength [ 10, 10] = "";
      // Loop through the requested cycles
      For (CycleI = CycleStart; CycleI <= CycleEnd; CycleI ++)
             // Reset the from and through numbers in the images from disk
             Swimming_T144 = ToText (CycleI);
             Swimming T145 = ToText (CycleI);
             // Clear overlaid screen objects
             ClearScreen ();
```

```
// Call the function which loads the images into memory
             Swimming_B120();
             // Play back the stack in memory
             For (ii = 0; ii < GetShape (Swimming_ArithmeticOpsNames) [0]; ii ++)
                    // Update user info
                    Swimming T118 =
                           "Processing Image Number":
                           ToText(ii):
                           " of ":
                           ToText (GetShape (Swimming_ArithmeticOpsNames) [0]):
              StatusBar = "Arithmetic Ops Copy of ": Swimming_ArithmeticOpsNames [ii,];
                    SelectROI ( Swimming_ROIImage );
                    ArithmeticOp ("Copy", Swimming_ArithmeticOpsNames [ii,]);
                    BeginOrEndUpdateBlock (TRUE);
                    SelectROI (Swimming ROIAnalyze):
                    Swimming\_Img = ii;
                    Swimming_SampleLoop();
                    BeginOrEndUpdateBlock (FALSE);
             // Loop through the rows
             For (j = 0; j < Swimming_NRows; j ++)
                    // Loop through the columns
                    For (i = 0; i < Swimming NCols; i ++)
                           MultipleMode = FALSE;
                           Local Real My_Points = Swimming_DataCentroid [i, j, 0...
Swimming_Img , 0 .. 2 ];
                           Temp_IDLine = CreateLine (My_Points);
                           If ( Temp_IDLine != FALSE )
                                  Extract ();
                                  Swimming_DataLength [i, j] = LnLength;
                                  Else
                                  Swimming_DataLength [ i, j] = 0.;
                           MultipleMode = TRUE;
             // If user opened data file send data to file
             If ( Swimming_cFileName )
```

```
// Open data file
Swimming_cFileHandle = OpenFile (Swimming_cFileName, 0x0001);
// Move to end of file
PositionFile (Swimming_cFileHandle, 0L, 2);
// Send data to data file
// -- Column Headings
If (CycleI == CycleStart)
       Swimming_cOutLine =
              "Col\t";
       // Loop through the rows
       For (j = 0; j < Swimming_NRows; j ++)
              // Loop through the columns
              For (i = 0; i < Swimming NCols; i ++)
                     Swimming cOutLine :=
                            ToText(i): "\t";
       Swimming cOutLine :=
              "\r\n" ;
       WriteFile (Swimming cFileHandle, Swimming cOutLine);
// -- Row Headings
If (CycleI == CycleStart)
       Swimming cOutLine =
              "Row\t";
       // Loop through the rows
       For (j = 0; j < Swimming_NRows; j ++)
              // Loop through the columns
              For (i = 0; i < Swimming_NCols; i ++)
                     Swimming_cOutLine :=
                            ToText ( j ): "\t";
       Swimming_cOutLine :=
              "\r\n";
       WriteFile (Swimming_cFileHandle, Swimming_cOutLine);
// -- Length Data
Swimming cOutLine =
       Swimming ArithmeticOpsNames [ Swimming Img , ] : "\t";
// Loop through the rows
For (j = 0; j < Swimming_NRows; j ++)
       // Loop through the columns
```

```
For (i = 0; i < Swimming_NCols; i ++ )
                                   Swimming_cOutLine :=
                                          ToText (Swimming_DataLength [i,j]): "\t";
                     Swimming cOutLine :=
                            "\r\n" ;
                     WriteFile (Swimming_cFileHandle, Swimming_cOutLine);
                     // Close data file
                     CloseFile (Swimming_cFileHandle);
              // Call the function which clears the images in memory
              Swimming_T118 = "Deleting Images from Stack!";
              Swimming_B122();
              }
       // Update user info
       Swimming_T118 = "Finished Processing Images!";
       }
// Button to prompt for data file info
Define Swimming B157 ()
       // Declare local variables
       Local Char TempPath [];
       // Initialize TempPath
       TempPath =
              FileSplitPath (ImageFile) [0,];
       TempPath :=
             FileSplitPath (ImageFile) [1,];
       // Prompt the user for the filename
       TempFileName = GetFileName (TempPath: "*.*", TRUE, 1, FALSE):
       // Get file name (with error trap)
       If ( TempFileName != FALSE )
              // Update the dialog box text
              Swimming_T161 = FileSplitPath (TempFileName) [0,];
              Swimming_T161 := FileSplitPath (TempFileName)[1,];
              Swimming_T162 = FileSplitPath (TempFileName) [2,];
              Swimming_T163 = FileSplitPath (TempFileName) [3,];
              Swimming_cFileName = TempFileName;
             // Open data file
              Swimming_cFileHandle = OpenFile (Swimming_cFileName, 0x0001);
             // Move to end of file
```

```
PositionFile (Swimming cFileHandle, 0L, 2);
             // Send data to data file
             // Close data file
             CloseFile ( Swimming cFileHandle );
      }
Define Swimming1_TerminateMacro ()
      // Unload the dialog box library
      LoadMacroLibrary(Swimming_hLib);
      // If OPTIMAS was iconic, bring it back
      If (Swimming_IconOPTIMAS)
             RunMacro (PathVariable: "macros/maximize.mac");
      // Wildcard delete of "Swimming .*"
      ObjectWildCardList ("Swimming_.*", 2);
      BeginOrEndUpdateBlock (FALSE);
// ------ LINK OBJECT creation -----
OBJECT_ID Swimming_Links[,];
Swimming_Links =
// Text
109 : ObjectID ( Swimming_T109 ) : 0 : 0 : 0 : 0
110 : ObjectID ( Swimming_T110 ) : 0 : 0 : 0 ::
118 : ObjectID ( Swimming_T118 ) : 0 : 0 : 0 ::
// Buttons
119:0:ObjectID (Swimming_B119):0:0::
120 : 0 : ObjectID ( Swimming_B120 ) : 0 : 0 ::
121:0: ObjectID (Swimming_B121):0:0::
122:0:ObjectID (Swimming B122):0:0::
123:0:ObjectID (Swimming_B123):0:0::
124:0: ObjectID (Swimming B124):0:0::
140:0: ObjectID (Swimming_B140):0:0::
157:0: ObjectID (Swimming_B157):0:0::
// Edit Boxes
141: ObjectID (Swimming T141): ObjectID (Swimming Update): ObjectID (
Swimming T141):0::
142 : ObjectID (Swimming T142) : ObjectID (Swimming Update) : ObjectID (
Swimming_T142):0::
143 : ObjectID (Swimming T143) : ObjectID (Swimming Update) : ObjectID (
Swimming T143):0::
144 : ObjectID ( Swimming T144 ) : ObjectID ( Swimming Update ) : ObjectID (
Swimming T144):0::
145 : ObjectID ( Swimming_T145 ) : ObjectID ( Swimming_Update ) : ObjectID (
Swimming_T145 ) : 0 ::
146 : ObjectID ( Swimming_T146 ) : ObjectID ( Swimming_Update ) : ObjectID (
Swimming_T146):0::
147 : ObjectID ( Swimming_T147 ) : ObjectID ( Swimming_Update ) : ObjectID (
Swimming T147):0::
```

```
148 : ObjectID ( Swimming_T148 ) : ObjectID ( Swimming_Update ) : ObjectID (
Swimming T148):0::
149 : ObjectID ( Swimming_T149 ) : ObjectID ( Swimming_Update ) : ObjectID (
Swimming_T149):0::
161 : ObjectID ( Swimming_T161 ) : ObjectID ( Swimming_Update ) : ObjectID (
Swimming_T161):0::
162 : ObjectID ( Swimming_T162 ) : ObjectID ( Swimming_Update ) : ObjectID (
Swimming_T162):0::
163 : ObjectID ( Swimming_T163 ) : ObjectID ( Swimming_Update ) : ObjectID (
Swimming_T163):0::
// Dummv
999:0:0:0:0;
// ----- Display the dialog box!!! ------
OPTCreateDialog (
      Swimming Directory: "Swimming.dlg", // Dialog template file, replace this!
      Swimming_Links, // Control Link
      Swimming_InitMacro, // Init Macro
      Swimming1_TerminateMacro, // Close Macro (note unique prefix!)
      hWndVideo, // Parent window handle
      Swimming hWndDlg, // handle to the new dialog
       . // Title bar object
       // hMenu
      );
```